# DIESTERS OF 2-(E-3-METHYLOXIRANYL)HYDROQUINONE FROM PIMPINELLA DIVERSIFOLIA

VASU DEV, C. S. MATHELA,\* A. B. MELKANI,\* NIEDRE M. POPE, NOEL S. STURM† and ALBERT T. BOTTINI†

Department of Chemistry, California State Polytechnic University, Pomona, CA 91768, U.S.A.; \*Kumaun University, Nainital 263002, India; †University of California, Davis, CA 95616, U.S.A.

(Received 29 September 1988)

Key Word Index—Pimpinella diversifolia; Apiaceae; 1,2,4-trisubstituted benzenes; <sup>1</sup>H NMR spectra.

Abstract—Three minor components of the essential oil from roots of *Pimpinella diversifolia*, gathered in the Kumaun Region of India, have been identified, primarily on the basis of their <sup>1</sup>H NMR spectra, as the mixed angelate-isobutyrate esters and the diangelate ester of 2-(E-3-methyloxiranyl)hydroquinone.

### INTRODUCTION

The (+)-Z-2-methyl-2-butenoate (angelate) and (+)-isobutyrate esters of 4-methoxy-2-(E-3-methyloxiranyl) phenol (1a and 1b) are the major constituents of the essential oil from roots of pre-flowering Pimpinella diversifolia DC. gathered in the Kumaun Region of India [1]. Examination of the oil by GC-MS showed the presence of three minor components with longer R<sub>t</sub>s than those of 1a and 1b and with relatively simple fragmentation patterns that indicated they were not terpenes. We describe here the isolation of these components and their identification as the previously unreported 1-angelyloxy-2-(E-3-methyloxiranyl)-4-isobutyryloxybenzene (2a), 1-isobutyryloxy-2-(E-3-methyloxiranyl)-4-angelyloxybenzene (2b) and 1,4-diangelyoxy-2-(E-3-methyloxiranyl)benzene (2c).

### RESULTS AND DISCUSSION

Compounds 2a—c were isolated as colourless oils in mg quantities by a modification of the chromatographic methods used to isolate 1a and 1b. The  $^1H$  NMR spectra of 2a—c are strikingly similar to those of 1a and 1b. Their signals at lowest field ( $\delta 6.95$ –7.11) show that the compounds are 1,4-trisubstituted benzenes [2], and their signals at  $\delta 3.60$  (J = 2.0 Hz), 2.92 or 2.93, and ca 1.4 show the presence of an E-3-methyloxiranyl group [3]. Signals at about  $\delta 6.3$  (1H) and between  $\delta 2.1$  and 2.0 (6H) characteristic of the angelyloxy part [4] are seen in the spectra of 2a and 2b, and two closely spaced sets of these signals (14H total) are seen in the spectrum of 2c. The spectra of 2a and 2b also have signals characteristic of the CHMe<sub>2</sub> part of an isobutyryl group. The gem-dimethyl

$$\mathbf{a} \quad \mathbf{R} = \mathbf{C}(\mathbf{Me}) = \mathbf{C}\mathbf{H}\mathbf{Me}$$

$$\mathbf{b} \quad \mathbf{R} = \mathbf{C}\mathbf{H}\mathbf{Me}_2$$

**a** 
$$R^1 = C(Me) = \frac{Z}{CHMe}$$
,  $R^2 = CHMe_2$   
**b**  $R^1 = CHMe_2$ ,  $R^2 = C(Me) = \frac{Z}{CHMe}$   
**c**  $R^1 = R^2 = C(Me) = \frac{Z}{CHMe}$ 

Short Reports

signal in the spectrum of 2a is a doublet at  $\delta 1.29$ . In the spectrum of 2b, as in that of 1b, the *gem*-dimethyl signal consists of two doublets at ca  $\delta 1.34$  separated by <0.01 ppm by the influence of one or both chiral centres of the 3-methyloxiranyl group [5], which must be *ortho* to the isobutyryloxy group.

The essential oil used for this study was from roots of plants gathered in the flowering stage rather than the preflowering stage. It was notably richer in 1a and 1b (54 and 21% compared with 37 and 15%) as well as 2a (0.8%), 2b (3.6%) and 2c (1.2%), but their relative amounts were similar

## **EXPERIMENTAL**

Isolation of **2a–c**. Pimpinella diversifolia in the flowering stage, was gathered in Sept. from the same location and the essential oil was obtained as described earlier. Compounds **2a–c** were obtained together by CC (silica gel, hexane–Et<sub>2</sub>O) of the essential oil and separated by HPLC on a  $10-\mu$  Porasil column using hexane–Et<sub>2</sub>O (17:3) as the mobile phase. From 2.42 g of the essential oil were obtained 1–3 mg of **2a–c** as colourless oils. The purity of the samples was >90% as determined by GC using a Varian 3700 GC equipped with a CDS 111 microprocessor, an FID and a 30-m J&W fused silica capillary column coated with DB-5.  $R_r$ s observed for **2a–c** were 70.4, 71.4 and 78.6 min under conditions that gave  $R_r$ s of 57.3 and 50.5 min for **1a** and **1b**.

1-Angelyloxy-2-(E-3-methyloxiranyl)-4-isobutyryloxybenzene (2a). HREIMS (probe) 70 eV, m/z (rel. int., BP 83) 318.1469  $[C_{18}H_{22}O_5]^+$  (8.4), 248.1031  $[C_{14}H_{16}O_4]^+$  (8.2), 236.1044  $[C_{13}H_{16}O_4]^+$  (54.6), 166.0635  $[C_9H_{10}O_3]^+$  (29.8), 148.0528  $[C_9H_8O_2]^+$  (4.10)  $[C_{18}H_{22}O_5]$  requires: 318.1467]. <sup>1</sup>H NMR (360 MHz, CDCl<sub>3</sub>): δ7.10 (d, J = 8.7 Hz, H-6), 7.00 (dd, J = 8.7, 2.7 Hz, H-5), 6.95 (d, J = 2.7 Hz, H-3), 6.30 (qq, J = 7.1, 1.6 Hz, H-3'), 3.60 (d, J = 2.0 Hz, H-1''), 2.93 (dd, J = 5.1, 2.0 Hz, H<sub>3</sub>-4'), 2.06 (app p, J = 1.6 Hz, Me-2'), 1.39 (d, J = 5.1 Hz, Me-3'') and 1.29 (d, J = 7.0 Hz, HCMe<sub>2</sub>).

1-Isobutyryloxy-2-(E-3-methyloxiranyl)-4-angelyloxybenzene (**2b**). HREIMS (probe) 70 eV, m/z (rel. int., BP 83) 318.1481  $[C_{18}H_{22}O_5]^+$  (11.5), 248.1020  $[C_{14}H_{16}O_4]^+$  (10.1), 236.1043  $[C_{13}H_{16}O_4]^+$  (59.8), 166.0639  $[C_9H_{10}O_3]^+$  (40.6), 148.0528  $[C_9H_8O_2]^+$  (49.7)  $[C_{18}H_{22}O_5]$  requires: 318.1467]. <sup>1</sup>H NMR

(360 MHz, CDCl<sub>3</sub>):  $\delta$ 7.05 (app d, J = 1.6 Hz, H-5 and H-6), 6.97 (app t,  $J_{app}$  = 1.6 Hz, H-3) 6.25 (qq, J = 7.0, 1.6 Hz, H-3'), 3.60 (d, J = 2.0 Hz, H-1"), 2.92 (q, d, J = 5.1, 2.0 Hz, H-3"), 2.86 (septet, J = 7.0 Hz, CHMe<sub>2</sub>), 2.06 (dq, J = 7.1, 1.4 Hz, H<sub>3</sub>-4'), 2.01 (m, Me-2'), 1.42 (d, J = 5.1 Hz, Me-3"), 1.335 (d, J = 7.0 Hz, MeCHMe) and 1.330 (d, J = 7.0 Hz, MeCHMe).

1,4-Diangelyloxy-2-(E-3-methyloxiranyl)benzene (2c). HREIMS (probe) 70 eV, m/z (rel. int., BP 83) 330.1469  $[C_{19}H_{22}O_5]^+$  (9.8), 248.1049  $[C_{14}H_{16}O_4]^+$  (45.3), 230.0943  $[C_{14}H_{14}O_3]^+$  (7.8), 166.0639  $[C_9H_{10}O_3]^+$  (3.1), 148.0515  $[C_9H_8O_2]^+$  (23.4)  $[C_{19}H_{22}O_5]$  requires: 330.1467]. <sup>1</sup>H NMR (360 MHz, CDCl<sub>3</sub>):  $\delta$ 7.11 (d, d = 8.7 Hz, H-6), 7.05 (dd, d = 8.7, 2.7 Hz, H-5), 6.99 (d, d = 2.7 Hz, H-3), 6.30 (d, d = 7.0, 1.6 Hz, H-3'), 6.24 (d, d = 7.0, 1.6 Hz, H-3''), 3.60 (d, d = 2.0 Hz, H-1''), 2.93 (d, d = 5.1, 2.0 Hz, H-3''), 2.09-2.02 (d-H<sub>3</sub>, H<sub>3</sub>-4''' Me-2', Me-2''') and 1.39 (d, d = 5.1 Hz, Me-3'').

Acknowledgements—This work was supported in part by a grant from the University Grants Commission, India, to CSM. High resolution mass spectra were obtained at the Mass Spectrometry Laboratory, Facility for Advanced Instrumentation, A. D. Jones (Director, University of California, Davis); the FAB-HS mass spectrometer, VG Analytical, used for determination of the spectra was purchased in part by NIH Division of Research Resources Grant RR01460. NMR spectra were obtained at the University of California, Davis, NMR Facility. We are grateful to Professor Freeman Allen (Pomona College, Claremont) for assistance in obtaining GC-MS data and the Council of Scientific and Industrial Research, India, for a fellowship to ABM.

### REFERENCES

- Bottini, A. T., Dev, V., Garfagnoli, D. J., Mathela, C. S., Melkani, A. B., Miller, A. A. and Sturm, N. S. (1986) Phytochemistry 25, 207.
- Silverstein, R. M., Bassler, G. C. and Morill, T. C. (1981) Spectrophotometric Identification of Organic Compounds, 4th Edn., Chap. 4. Wiley, New York.
- 3. Villa, L., Schenetti, L. and Taddei, F. (1973) Org. Magn. Reson. 5, 593.
- Brouwer, C. H. and Stothers, J. B. (1972) Can. J. Chem. 50, 601.
- Whitesides, G. M., Holtz, D. and Roberts, J. D. (1964) J. Am. Chem. Soc. 86, 2628.